

1. Reversed Phase Specialty Columns

Specifications

Packing Material	Cholester		PBr		PFP	π NAP	PYE	NPE		
Silica Gel*	Fully-Porous		Core-Shell	Fully-Porous	Core-Shell	Fully-Porous	Fully-Porous	Fully-Porous		
Average Particle Size (μm)	2.5	3	5	2.6	3	5	2.5	3	5	
Average Pore Size (\AA)	130	120	90	120	90	120	130	120	120	
Specific Surface Area (m^2/g)	330	300	150	300	150	300	330	300	300	
Bonded Phase Structure										
Bonded Phase	Cholesteryl group		Pentabromobenzyl group	Pentafluorophenyl group	Naphthylethyl group	Pyrenylethyl group	Nitrophenylethyl group			
Bonding Type	Monomeric									
Main Interaction	Hydrophobic interaction Molecular shape selectivity		Hydrophobic interaction Dispersion force		Hydrophobic interaction π - π interaction Dipole-dipole interaction	Hydrophobic interaction π - π interaction	Hydrophobic interaction π - π interaction Dispersion force Molecular shape selectivity	Hydrophobic interaction π - π interaction Dispersion force Dipole-dipole interaction		
End-Capping	Near-perfect treatment									
Carbon Content	21%	20%	—	8%	—	10%	14%	11%	18%	9%
Usable pH Range	2 ~ 7.5									
Features	<ul style="list-style-type: none"> Usable under the same conditions as C₁₈ High molecular shape selectivity 		<ul style="list-style-type: none"> Separate hydrophilic compounds under reversed-phase conditions Separate using dispersion force 		<ul style="list-style-type: none"> Weak dipole-dipole interaction 	<ul style="list-style-type: none"> Stronger π-π interaction than phenyl column 	<ul style="list-style-type: none"> Very strong π-π interaction 	<ul style="list-style-type: none"> Strong dipole-dipole interaction 		

* Silica Gel : Fully-Porous···High purity porous spherical silica Core-Shell···Core-Shell silica gel

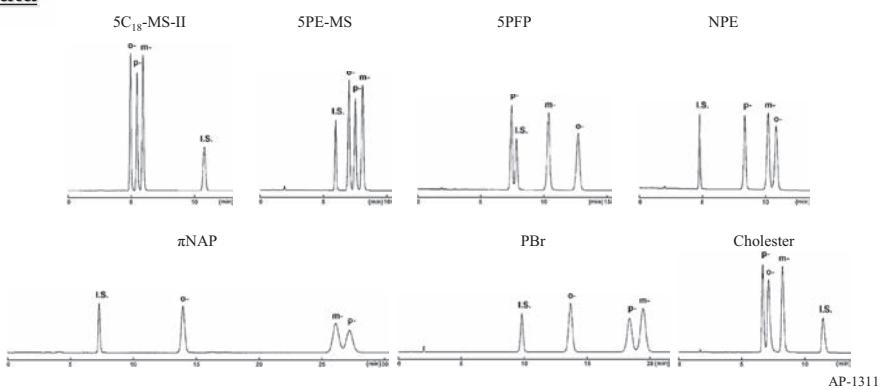
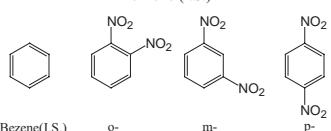
Selectivity for positional isomers of dinitrobenzene

Different stationary phases exhibit different selectivity due to the use of forces that C₁₈ (hydrophobic interaction) does not have. By using these columns, you can achieve separation that cannot be done using only C₁₈.

COSMOSIL Application Data

Column: COSMOSIL **
 Column size: 4.6mmI.D.-150mm
 Mobile phase: Methanol/ H₂O = 50/50
 Flow rate: 1.0 ml/min
 Temperature: 30°C
 Detection: UV254nm

Sample: o-Dinitrobenzene
 m-Dinitrobenzene
 p-Dinitrobenzene
 Benzene (I.S.)



AP-1311



COSMOSIL Cholester / COSMOCORE Cholester

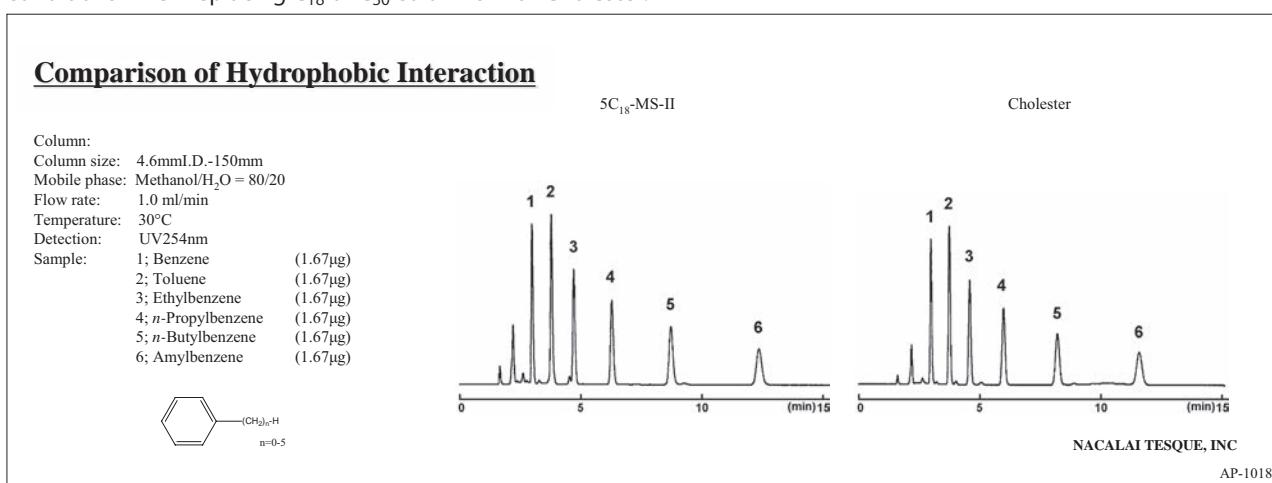
- Cholesterol-bonded stationary phase
- Increased stereoselectivity and improved resolution for geometric isomers
- Usable under the same conditions as C₁₈

Suitable Samples

- Natural compounds
- Structurally similar compounds
- Polyphenols, catechins, fat-soluble vitamins and flavones

Hydrophobic Interaction

The below figure shows the comparison of hydrophobic interactions with competitor C₁₈ columns. Cholester provides about the same hydrophobicity as alkyl group-bonded types (C₁₈, C₃₀). It is not necessary to change the analytical conditions when replacing C₁₈ or C₃₀ columns with Cholester.



Molecular Shape Selectivity

The stationary phase of Cholester has a very rigid structure and can distinguish different molecular shapes. Cholester retains planar triphenylene longer than non-planar o-Terphenyl.

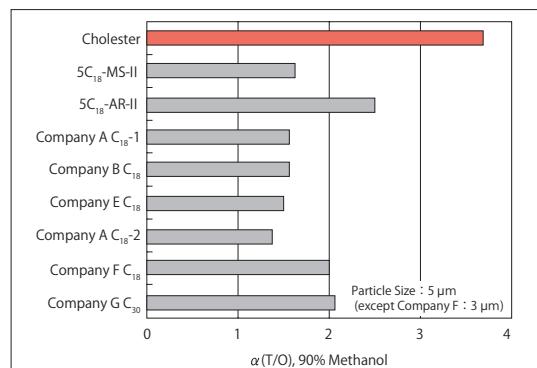
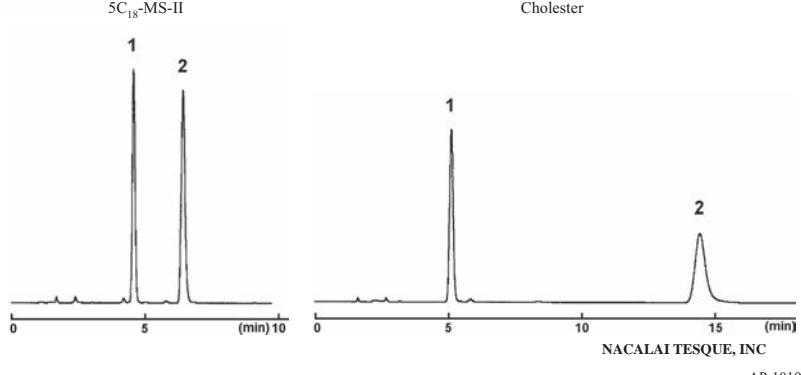
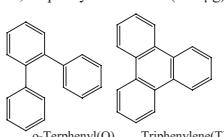


Figure. Comparison of molecular shape selectivity

Comparison of Molecular Shape Selectivity

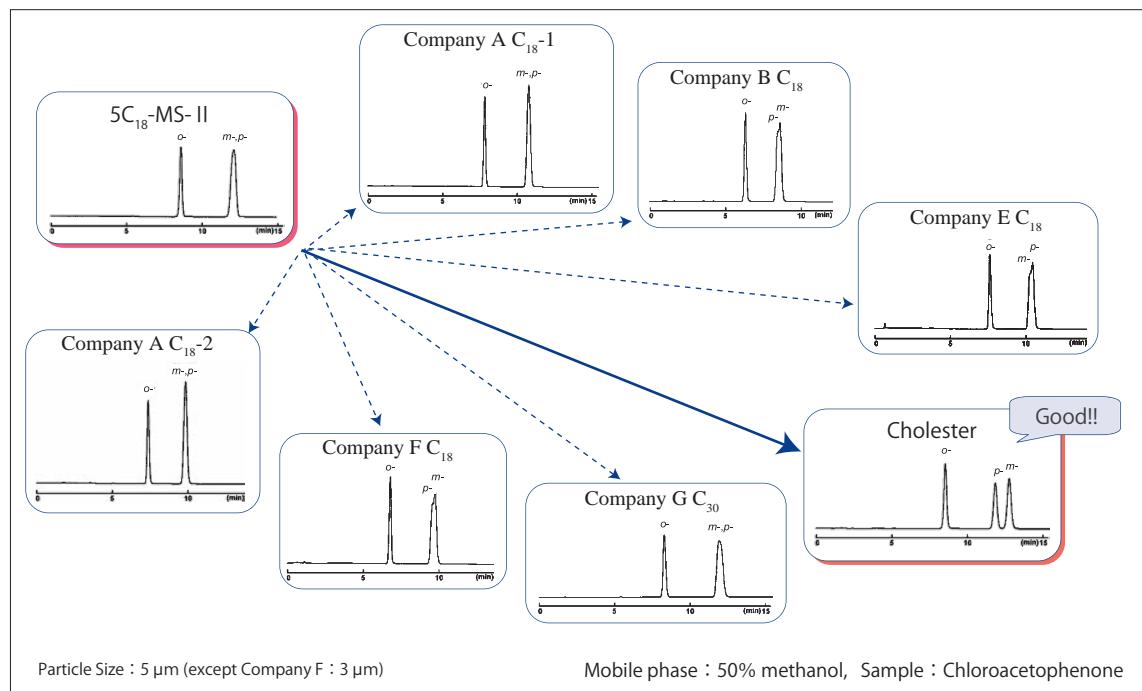
- Column:
Column size: 4.6mmI.D.-150mm
Mobile phase: Methanol/H₂O = 90/10
Flow rate: 1.0 ml/min
Temperature: 30°C
Detection: UV254nm
- Sample:
1; o-Terphenyl (0.1μg)
2; Triphenylene (0.01μg)



Improvement in Separation

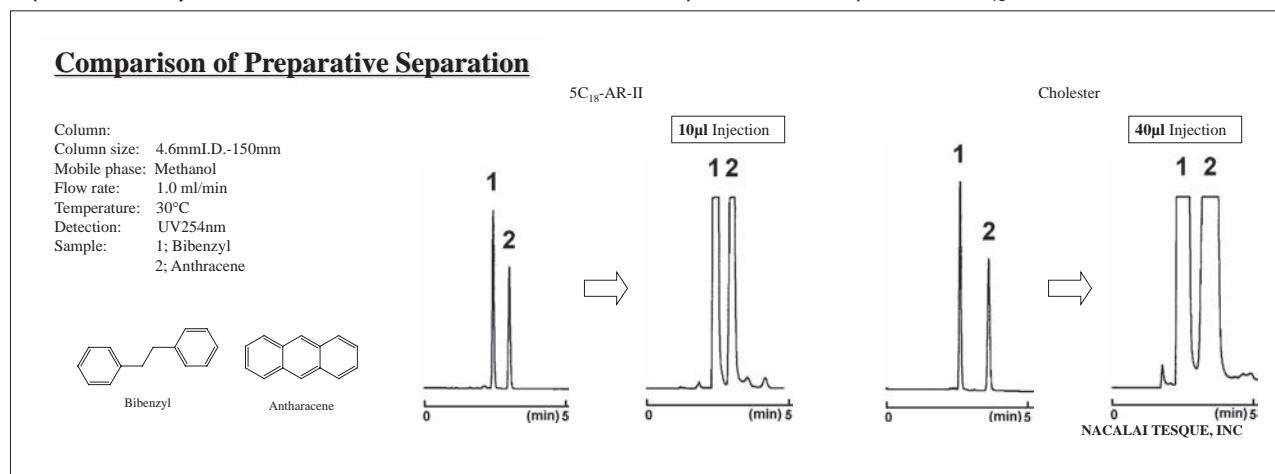
COSMOSIL Cholester provides enhanced selectivity over traditional C₁₈ columns and offers greater performance in separating isomers or other closely related compounds. COSMOSIL Cholester is ideal for method development and serves as an excellent alternative to traditional C₁₈ columns. The figure below shows analytical data of chloroacetophenone isomers. These isomers are difficult to separate with C₁₈ and C₃₀, but they are well resolved by COSMOSIL Cholester.

● Comparison with competitor's C₁₈ and C₃₀ columns



Efficiency of Preparative Separation

The figure below shows the comparison of efficiency of preparative separation with a C₁₈ column. Both columns show good separation. However, sample loading capacity for preparative separations can be affected by a slight difference in separation ability. COSMOSIL Cholester can load 4 times the sample volume compared with C₁₈ columns.



About Core-Shell 2.6Cholester Particles

COSMOCORE 2.6Cholester is packed with cholesterol-bonded 2.6 μm core-shell particles. It delivers performance equivalent to sub-2 μm particles at faster flow rate and analysis time while maintaining a lower back pressure. COSMOCORE can also be used in longer column size to gain additional resolution.

● Catechins (Standard)

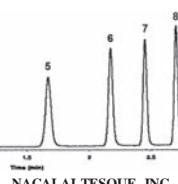
COSMOCORE Application Data

Column: COSMOCORE 2.6Cholester
Column size: 2.1mmL.D.-50mm
Mobile phase: A; Acetonitrile/ 0.1% H_3PO_4 = 10/90
B; Acetonitrile/ 0.1% H_3PO_4 = 40/60
B conc. 0 → 100% 3min Linear gradient

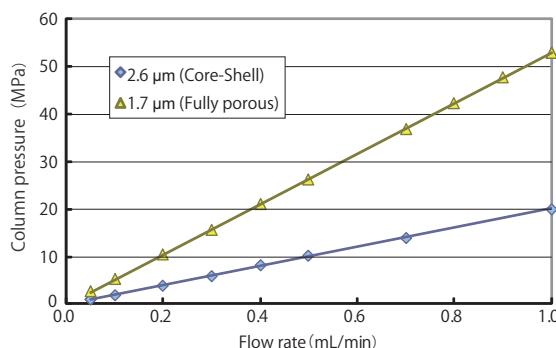
Flow rate: 0.6 mL/min
Temperature: 30°C
Detection: UV280nm

Sample:
1: (-)-Gallocatechin[(-)-GC]
2: Caffeine
3: (-)-Epigallocatechin[(-)-EGC]
4: (-)-Catechin[(-)-C]
5: (-)-Epicatechin[(-)-EC]
6: (-)-Epigallocatechin Gallate[(-)-EGCg]
7: (-)-Gallocatechin Gallate[(-)-GCG]
8: (-)-Epicatechin Gallate[(-)-ECg]
9: (-)-Catechin Gallate[(-)-Cg]

Data courtesy of a customer



● Comparison of Column Pressure



Column Size : 2.1 mmL.D. x 100 mm
Mobile Phase : Acetonitrile/Water = 70/30
Temperature : 40°C

Applications

● Catechins (Commercial Green Tea)

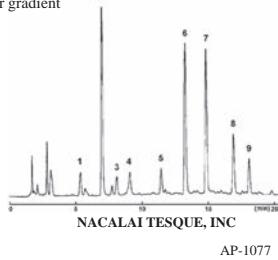
COSMOSIL Application Data

Column: COSMOSIL Cholester
Column size: 4.6mmL.D.-150mm
Mobile phase: A; Acetonitrile/ 20mmol/l Phosphate buffer(pH2.5) = 10/90
B; Acetonitrile/ 20mmol/l Phosphate buffer(pH2.5) = 30/70
B conc. 0 100% 20min Linear gradient

Flow rate: 1.0 mL/min
Temperature: 30°C
Detection: UV280nm

Sample: Green Tea
1: (-)- Galloatechin [(-)- GC]
2: Caffeine
3: (-)- Epigalloatechin [(-)- EGC]
4: (-)- Catechin [(-)- C]
5: (-)- Epicatechin [(-)- EC]
6: (-)- Epigallocatechin Gallate [(-)- EGCG]
7: (-)- Gallocatechin Gallate [(-)- GCG]
8: (-)- Epicatechin Gallate [(-)- ECg]
9: (-)- Catechin Gallate [(-)- Cg]

Injection Vol. 1.0μl

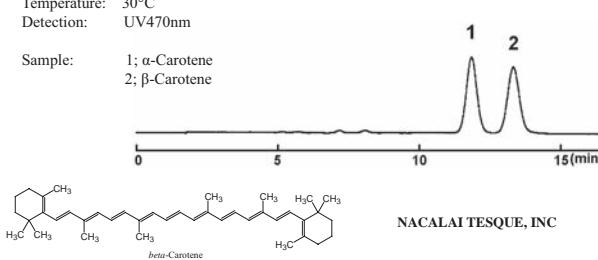


● Carotene

COSMOSIL Application Data

Column: COSMOSIL Cholester
Column size: 4.6mmL.D.-150mm
Mobile phase: Tetrahydrofuran/Methanol = 20/80
Flow rate: 1.0 mL/min
Temperature: 30°C
Detection: UV470nm

Sample: 1; α -Carotene
2; β -Carotene



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● Fat-Soluble Vitamins

COSMOSIL Application Data

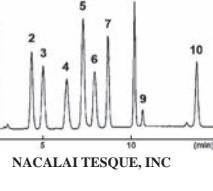
Column: COSMOSIL 2.5Cholester
Mobile phase: A; 0.1% TFA-Methanol/ H_2O = 90/10
B; 0.1% TFA-Methanol
B conc. 0% (0-5min)-100%(10min)-100%(15min)

Flow rate: 0.4 mL/min
Temperature: 40°C
Detection: UV280nm

Sample:

1: Vitamin A Acetate, *all trans* (0.06mg/ml)
2: Vitamin D₂ [Calciferol] (0.30mg/ml)
3: Vitamin D₃ (0.06mg/ml)
4: Vitamin E Succinate [D- -Tocopherol Succinate] (0.60mg/ml)
5: Vitamin K₂ (0.18mg/ml)
6: Vitamin E [DL- -Tocopherol] (0.60mg/ml)
7: Vitamin E Acetate [DL- -Tocopherol Acetate] (0.60mg/ml)
8: *trans*-Vitamin K₁ (0.18mg/ml)
9: *cis*-Vitamin K₁
10: Vitamin A Palmitate (0.18mg/ml)

Inj. Vol: 2.5μl



● Flavone

COSMOSIL Application Data

Column: COSMOSIL Cholester
Column size: 4.6mmL.D.-150mm
Mobile phase: A: Acetonitrile/ 20mmol/l Phosphate buffer(pH2.5) = 20/80
B: Acetonitrile/ 20mmol/l Phosphate buffer(pH2.5) = 70/30
B conc. 0 100% 20min Linear gradient

Flow rate: 1.0 mL/min
Temperature: 30°C
Detection: UV280nm

Sample:

1: Fisetin (0.25μg)
2: Myricetin (0.20μg)
3: 7,8-Dihydroxyflavone (0.05μg)
4: Luteolin (0.20μg)
5: Quercetin (0.20μg)
6: 7-Hydroxyflavone (0.10μg)
7: Baicalein (0.05μg)
8: 6-Hydroxyflavone (0.05μg)
9: Flavon (0.05μg)
10: Chrysin (0.05μg)
11: 6-Methoxyflavone (0.05μg)
12: 3-Hydroxyflavone (0.25μg)
13: 5-Hydroxyflavone (0.05μg)

NACALAI TESQUE, INC

